Cystic fibrosis: current concepts

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Abstract

Cystic fibrosis is an autosomal recessive genetic disease, mainly in Caucasian children and young adults. It is caused by pathogenic variants in the *CFTR* (cystic fibrosis transmembrane conductance regulator) gene, which results in increased viscosity and difficult mucus clearance. The main organ affected is the lung, the pancreas, sweat glands, intestine, liver, nasal mucosa, salivary glands, and reproductive tract. The clinical manifestations vary, ranging from the most frequent pulmonary symptoms of obstructive disease to gastrointestinal manifestations related to malabsorption secondary to pancreatic insufficiency. Although there are multiple diagnostic tests for cystic fibrosis, neonatal screening to identify increased immunoreactive trypsinogen, chloride sweat test, and the detection of pathogenic variants in the *CFTR* gene allow the diagnosis to be integrated. Cystic fibrosis management consists of three main strategies: firstly, to keep the airway free of secretion; secondly, to keep the airway free of infection; and finally, to maintain an optimal nutritional status. Therapies that seek to correct alterations in the *CFTR* gene are focused on avoiding a pathogenic nonsense variant, correcting folding, increasing trafficking to the plasma membrane, or increasing the function of the *CFTR* channel. Other therapies still under development include gene therapy, genome editing, and antisense oligonucleotides to modify the expression of this gene.

Keywords: Cystic fibrosis. *CFTR* gene. Gene therapy.

Fibrosis quística: conceptos actuales

Resumen

La fibrosis quística es una enfermedad genética autosómica recesiva que se presenta principalmente en niños y adultos jóvenes caucásicos. Está causada por variantes patogénicas en el gen *CFTR* (regulador de la conductancia transmembrana de la fibrosis quística), lo que ocasiona un aumento de la viscosidad y un difícil aclaramiento del moco. El principal órgano afectado es el pulmón, seguido del páncreas, las glándulas sudoríparas, el intestino, el hígado, la mucosa nasal, las glándulas salivales y el aparato reproductor. Las manifestaciones clínicas son variables y van desde las más frecuentes, que son los síntomas pulmonares de enfermedad obstructiva, hasta manifestaciones gastrointestinales relacionadas con la malabsorción secundaria a la insuficiencia pancreática. Aunque existen múltiples pruebas diagnósticas para la fibrosis quística, el tamiz neonatal, el aumento en el tripsinógeno inmunorreactivo, la prueba de cloro en sudor y la detección de variantes patogénicas en el gen *CFTR* permiten integrar el diagnóstico. El manejo consta de tres niveles principales: el primero, mantener la vía aérea libre de secreción; el segundo, mantener la vía aérea libre de infección; y por último, mantener un estado nutricional óptimo. Las terapias que buscan corregir las alteraciones en el gen *CFTR* están enfocadas en evitar una variante

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Date of reception: 16-11-2020

Date of acceptance: 03-05-2021

DOI: 10.24875/BMHIM.20000372

Available online: 16-12-2021

Bol Med Hosp Infant Mex. 2021;78(6):584-596

www.bmhim.com

patogénica sin sentido, corregir el plegamiento, aumentar el tráfico a la membrana plasmática o incrementar la función del canal CFTR. Otras terapias aún en desarrollo incluyen la terapia génica, la edición del genoma y los oligonucleótidos antisentido para modificar la expresión de este gen.

Palabras clave: Fibrosis quística. Gen CFTR. Terapia génica.

Introduction

Cystic fibrosis (CF), a monogenic disease transmitted in an autosomal recessive pattern, is multisystemic and chronic and originates as a consequence of pathogenic changes in the *CFTR* gene located on the long arm of chromosome 7 (locus 7q.31) that encodes for the protein known as the CF transmembrane conductance regulator (CFTR). Dysfunction of this protein causes altered ion transport in the apical membrane of epithelial cells in different organs. This dysfunction becomes evident in the early stages of life (infant stage). It is a complex and highly pleomorphic disease whose classic phenotype is a progressive obstructive pulmonary disease, exocrine pancreatic insufficiency, and elevated sweat chloride and sodium levels in 90% of patients¹.

Epidemiology

CF is the most common inherited genetic disease among Caucasians; however, any ethnic group can be affected. Prevalence ranges from 1:1400 in Ireland to 1:3500 in the United States, while rates are much lower in geographic regions such as Asia and Africa^{2,3}. In Latin America, the prevalence is 1:1600-14,000 live newborns (LNB), and in Mexico, 1:8500 LNB, with a detection rate of pathogenic variants of 41.6%^{4,5}.

Mortality in childhood is due to complications of recurrent respiratory infections and malnutrition due to pancreatic insufficiency⁶. However, in Mexico and Latin America, life expectancy in the early 1990s averaged only 9 years. With the use of new therapies and better control of the disease, the average survival is 18 years¹. In developed countries, life expectancy exceeds 40 years; these "long survivors" have a mean survival of 52.1 years of age in Canada, 47.4 years in the United States, and 47.3 years in the United Kingdom^{1,7}.

Etiopathogenesis

CF is caused by pathogenic variants in the *CFTR* gene (locus 7q31.2) and is inherited in an autosomal recessive (AR) manner⁸⁻¹⁰. More than 2000 variants have been identified in the *CFTR* gene, but the most common variant is the phenylalanine deletion at

position 508 (p.F508del, Δ F508)^{11,12}. The pathogenic variants and their effects have been grouped into seven classes (Table 1)¹²⁻¹⁴.

Structurally, CFTR is a membrane-bound glycoprotein with a typical 12-helix architecture comprising two pseudosymmetric transmembrane domains (MSD) and two nuclear binding domains (NBD) that bind and hydrolyze ATP. Between both NBD units is a single regulatory domain (R) consisting of many charged amino acids (Figure 1)¹⁵. The CFTR protein is expressed mainly in the lung and the pancreas, sweat glands, intestine, liver, nasal mucosa, salivary glands, and reproductive tract¹⁶. Under normal conditions, CFTR works like a gate tightly coupled to ATPase cycles (Figure 1) through NBD dimerization, which subsequently induces the formation of a transmembrane domain cavity that opens towards the extracellular side to allow the flow of selective anions as chlorine (Cl-) and bicarbonate (HCO₃-)¹¹.

Particularly in the airway, CFTR regulates the local pH by allowing Cl- and HCO₃- to flow out of the cell; in addition, it causes the epithelial sodium channel ENaC to transport sodium into the cell (Figure 2). Normal airway surface fluid (ASF) has a pH around 7.0¹⁷. However, in CF, the pH is eight times more acidic due to the lack of adequate secretion of Cl- and HCO₃- anions into the extracellular space (Figure 2). The loss of chloride secretion due to CFTR deficiency results in changes in osmotic pressures and electroneutrality, which probably lead to excessive sodium and water absorption¹⁸.

Pulmonary alterations are due to the addition of a considerably more acidic pH (caused by the lack of secretion of Cl- and HCO₃- anions) and the excessive absorption of Na+ and water that cause an increase in mucus viscosity and a difficult clearance of mucus in the airway. In the other organs and systems, CF produces obstructions at different levels, causing extrapulmonary symptoms.

Clinical manifestations

Respiratory, gastrointestinal, and metabolic alterations are present in CF, some associated with inadequate nutrient uptake related to intestinal pathology.

Table 1. Classification of *CFTR* gene mutations. The most common mutations in each class and the type of defect are described

Classification of CFTR mutations	Description	Examples
<p>Normal</p>	<p>The CFTR protein is found on the cell surface and functions properly, allowing chloride and water transfer</p>	—
<p>Class I</p>	<p>No protein production: No CFTR protein synthesis</p>	<p>p.G542X p.W1282X p.R553X</p>
<p>Class II</p>	<p>Traffic defects: CFTR protein is created but is misfolded, preventing it from reaching the cell surface</p>	<p>p.F508del p.N1303K p.I507del</p>
<p>Class III</p>	<p>Defective regulation: CFTR protein is created and reaches the cell surface, but there is a defect in the channel opening</p>	<p>p.G551D p.S549N p.V520F</p>
<p>Class IV</p>	<p>Decreased conductance: There is a defect in ion transport through the CFTR protein, resulting in a poor outflow of ions from the cell</p>	<p>p.R117H p.D1152H p.R347P</p>
<p>Class V</p>	<p>Reduced amount: CFTR protein is created but in insufficient quantities, resulting in decreased ion transport</p>	<p>c. 621+1G>T c. 3849+10C>T c. 2780+5G>A p.A455E c. 3140-26A>G</p>
<p>Class VI</p>	<p>Increased turnover: A decrease in the half-life of the CFTR protein occurs, causing accelerated turnover of the CFTR protein</p>	<p>c. 4326delTC p.Gln1412X</p>
<p>Class VII</p>	<p>No mRNA: No mRNA is synthesized, no CFTR protein is formed</p>	<p>Dele2,3 (21kb)</p>

CFTR, cystic fibrosis transmembrane conductance regulator; mRNA, messenger ribonucleic acid.

Figure 1. Representation of the CFTR protein channel, composed of two transmembrane domains (MSD 1 and 2), each bound to a nucleotide-binding domain (NBD 1 and 2). NBD1 is connected to MSD2 by a regulatory domain (R). NBD 2 has a carboxyl end. CFTR, CF transmembrane conductance regulator; MSD, transmembrane domain; NBD, nucleotide-binding domain; R, regulatory domain.

Figure 2. The function of the CFTR channel, the function of the ENaC channel, and mucin secretion for the formation of normal airway surface fluid are illustrated. **A.** Cell dynamics in a healthy context. **B.** Cell dynamics in CF in the context of the p.F508del mutation. The intradomain defect affects full-length CFTR protein assembly and post-translational stability, whereby deletion of phenylalanine at position 508 in NBD1 leads to a CFTR trafficking defect. CFTR, cystic fibrosis transmembrane conductance regulator; ENaC, epithelial sodium channel.

The presenting symptoms may vary according to genotype. Prior to the implementation of newborn age at diagnosis and are sometimes related to screening in the United States, the diagnosis of CF in

infants and preschoolers was integrated after presenting with one or more of the following symptoms¹⁹: respiratory symptoms (45% of patients), inadequate weight for age (28%), or meconium ileus (20%). In addition, the clinical presentation in adults with atypical features is better in the long term than those diagnosed in childhood¹⁹.

Signs and symptoms can be grouped according to the age group or location of the damage. According to age group, meconium ileus, prolonged neonatal jaundice (cholestatic), edema, anemia, malnutrition, steatorrhea, malabsorption syndrome, inadequate weight gain, and recurrent vomiting may be found in newborns and young infants. There is often recurrent or chronic coughing or wheezing in infants that do not improve with treatment, recurrent or chronic pneumonia, growth retardation, chronic diarrhea, rectal prolapse, salty skin taste, and chronic hyponatremia, and hypochloremia. Preschool children may show chronic cough with purulent expectoration and chronic recurrent unexplained wheezing (both with no response to treatment), poor weight and height gain, recurrent abdominal pain, rectal prolapse, intussusception, chronic diarrhea, clubbing of the fingers, chronic hyponatremia, and chronic hypochloremia, hepatomegaly or unexplained liver disease, and nasal polyps. Schoolchildren present with chronic unexplained respiratory symptoms, *Pseudomonas aeruginosa* in bronchial discharge, chronic sinusitis, nasal polypsis, bronchiectasis, chronic diarrhea, distal intestinal obstruction syndrome, pancreatitis, rectal prolapse, and hepatomegaly. Moreover, adolescents and adults often present with chronic unexplained suppurative lung disease, digital clubbing, recurrent abdominal pain, pancreatitis, distal intestinal obstruction syndrome, liver cirrhosis and portal hypertension, growth retardation, male sterility with azoospermia, and decreased fertility in women²⁰. These symptoms are characteristic in patients with chloride concentrations > 60 mmol/L (discussed later). However, a subgroup of patients with chloride concentrations in the intermediate range is diagnostically challenging. As mentioned by Jorquera²¹, these patients may develop minimal symptoms or other symptoms related to defects in the *CFTR* gene in non-respiratory organs, making diagnosis difficult. Symptoms usually range from clubbing toes, recurrent respiratory infections, and isolation of *P. aeruginosa* and *Staphylococcus aureus*. Less common symptoms are pancreatic insufficiency, meconium ileus, distal intestinal obstruction syndrome, and liver disease²¹.

Respiratory symptoms

Respiratory alterations can be found in 80% of infants and preschoolers¹⁹. In children under one year of age, dry and repetitive cough is frequent. Also, persistent tachypnea, a slight increase in the anteroposterior diameter of the thorax, decreased expansion of the upper thorax, persistent intercostal retraction, and bronchial obstruction may be observed. Older children may present with bronchial obstruction and cough with purulent mucous secretions as constant symptoms. There are variable degrees of thoracic deformity with increased anteroposterior diameter. In the thoracic auscultation, in general, there is no added noise in inter-crisis periods, and crackles in infectious exacerbations are present in children with severe compromise. In advanced disease, bronchiectasis, with or without hemoptysis, watch glass nails, and clubbing fingers may be present, while cyanosis is a late sign²⁰.

Sinus disease is also present in most patients and manifests as chronic nasal congestion, headache, cough, chronic posterior nasal discharge, and sleep disorders¹⁹.

With increased viscosity and difficult clearance, respiratory secretions obstruct the airways and promote infection, leading to tissue destruction and, eventually, bronchiectasis. In chronic airway obstruction, *Haemophilus influenzae*, *S. aureus*, *P. aeruginosa*, and *Burkholderia cepacia* are the main pathogenic bacteria²².

Some germs have been implicated in the infection that occurs in CF. Before the antimicrobial era, few children with CF survived beyond the age of 2 years. Children died primarily from failure to thrive, along with bacterial pneumonia. The organism found in the lungs at autopsy in these children was almost always *S. aureus*, which remains the predominant pathogen of CF from birth to adolescence, with 80% of children infected in preadolescence and early adolescence. Moreover, the critical pathogen in chronic lung disease is an unusual morphotype of *P. aeruginosa* termed mucoid. Although this germ is comparatively avirulent, it induces a chronic inflammatory response, apparently responsible for the lung damage that leads to the patient's demise. This process may occur over years or decades, which would explain why 60-75% of adults in the United States are infected with this organism. In other cases of CF, a double count of glucose non-fermenting bacilli that are *B. cepacia* complex organisms, *Achromobacter* and *Stenotrophomonas*, are more frequently found in the patient. The latter is found in

approximately 10% to 20% of CF patients, while *Achromobacter* is found in 5% to 10%. The incidence of these organisms increases with age and probably reflects patients who have received multiple rounds of antipseudomonal antimicrobial therapy²³.

In the case of *P. aeruginosa*, responsible for morbidity and mortality in CF, a prevalence of 25-30% has been reported in infants aged 0-1 year, 38-40% in children aged 2-6 years, and can reach 80% in patients > 18 years. Infection is mainly caused by rough strains with high sensitivity to antibiotics. However, in the case of CF, this infection is not completed and acquires a mucoid phenotype in response to alteration of the *CFTR* gene epithelium. Therefore, the stagnation of mucus due to dehydration helps form an ideal culture medium and a layer around the bacteria that makes the immune system and antibiotics less effective. Consequently, endobronchial infection becomes chronic and can no longer be eliminated²⁴.

The most frequently detected symptoms in the more advanced stages are pulmonary hypertension, bronchiectasis, and *cor pulmonale*. Progressive lung disease remains the leading cause of morbidity and mortality in these patients²².

Signs and symptoms that evidence pulmonary exacerbation are increased cough, changes in sputum (consistency or volume), decreased appetite, or weight loss. Exacerbation also shows changes in respiratory examination and respiratory rate. However, the number and variability of signs and symptoms may alter the exacerbation pattern²⁵.

Extra-respiratory symptoms

Digestive system and nutrition. About 3.9% of patients do not present extrapulmonary manifestations related to pancreatic, gastrointestinal, and hepatobiliary dysfunction^{26,27}. Defects in the *CFTR* gene result in abnormalities affecting organs such as the liver, gastrointestinal tract, and pancreas. Nutritional failure in CF is multifactorial. Malabsorption of lipids, protein, and fat-soluble vitamins results from insufficient pancreatic enzyme production, which bile salt abnormalities may aggravate in the presence of concurrent liver disease. Because of the predisposition to pulmonary infection, a relationship has been found with increased breathing, reduced appetite, and increased calories at the expense of inflammatory catabolism. Approximately 85% of patients develop pancreatic insufficiency between 1-2 years of age. In the context of CF, pancreatic

insufficiency is a risk factor for recurrent pancreatitis, which is the presentation for the diagnosis of CF²⁶.

Gastrointestinal tract manifestations are related to mucosal dysmotility and include meconium ileus, constipation, distal intestinal obstruction syndrome, gastroesophageal reflux disease, and small intestinal bacterial overgrowth. The hepatobiliary disease has a prevalence of 10-15%, with peak onset in preadolescence.

In exclusively breastfed infants, the triad of anemia, hypoalbuminemia, and edema—secondary to malnutrition—and some specific deficiencies due to malabsorption caused by CF are common²⁰.

CF-related liver disease. Approximately 20-40% of people with CF develop clinically detectable CF-related liver disease (CFLD)^{19,26}.

Hypovitaminosis. CFLD and pancreatic dysfunction lead to lipid malabsorption, predisposing to deficiencies of fat-soluble vitamins (A, D, E, and K)²⁸.

CF-related diabetes (CFRD). The main cause is an insulin deficiency related to the destruction of pancreatic islets²⁹. Two percent of CF patients have CFRD in childhood, increasing to 20% of adolescents and reaching 40-50% of CF patients in adulthood. This increasing proportion suggests multifactorial, complex, and progressive pathogenesis of CFRD. Excessive protein catabolism is problematic in CF lungs, associated with an imbalance between catabolic and anabolic enzymes.

Studies in adults have shown that insulin depletion contributes to an inflammatory catabolic state, compromising lung function³⁰. It has been suggested that CFRD patients with no impaired basal glycemia do not require treatment because they are asymptomatic. However, the nutritional consequences of insulin deficiency may impair their quality of life. In the case of patients with altered basal glycemia, microalbuminuria was observed in 14%, retinopathy in 16%, neuropathy in 55%, and gastropathy in 50%. Therefore, from the age of 5 years, it is advisable to monitor annually for microvascular complications from the diagnosis of CFRD³¹.

Recurrent venous thrombosis. CF seems to be a risk factor for the development of recurrent venous thrombosis. Of 120 children and young adults with acute venous thromboembolism, 19 showed recurrent thrombosis. Among them, six had CF, of which five were infected with *B. cepacia*³².

Anemia. It occurs in 10% of children but is more frequent in older adults or those with decreased pulmonary function. Although its causes are diverse, the relationship with CF remains unclear, especially in infants, in whom it may occasionally be a presenting sign. Mechanisms involved include dysregulation of

iron metabolism, anemia due to chronic disease, blood loss, renal failure, or bone marrow suppression after transplantation²⁸.

Bone disease. Bone disease, characterized by decreased mineral density and increased rates of fracture and kyphosis, is rare in children under 10 years of age but increases in prevalence with age. In addition, vitamin D deficiency decreases intestinal absorption of calcium and produces secondary hyperparathyroidism, which leads to increased bone resorption and fragility¹.

Infertility. Most male CF patients (> 98%) have a congenital bilateral absence of the *vas deferens* (CBAVD) and are infertile, but < 2% remain fertile and usually carry CFTR genotypes. It has been proposed that reduced CFTR activity impairs ion exchange within the male genital ducts and predisposes to increased intraluminal viscosity and obstruction by accumulated secretions and mucus, ultimately leading to obliteration of epididymis and *vas deferens*^{33,34}. In contrast, women with CF experience more pulmonary exacerbations after puberty than men and increased colonization with the mucoid form of the commonly acquired CF pathogen *P. aeruginosa*. Gonadal hormones may contribute to these sex differences. Female sex hormones (estrogen and progesterone) are expressed in lung tissue and their hormone receptors. There is a 35% subfertility and infertility rate in women with CF trying to conceive (mean age 30.4 years). In the cervix, the defective CFTR in the epithelium promotes the production of thick, dehydrated cervical mucus. This mucus blocks the cervix and impairs the entry of spermatozoa. In the endometrium, defective CFTR in the epithelium can alter bicarbonate secretion, reducing uterine fluid volume and impairing sperm fertilization capacity. Finally, women with CF may have a lower ovarian reserve compared to age-matched controls³⁵.

Nephrolithiasis and nephrocalcinosis. CF has been associated with nephrolithiasis and nephrocalcinosis due to factors that promote calcium deposition.

Diagnosis

New challenges have emerged in both the diagnosis and clinical management of children with CF, as various forms of terminology for the disease have been introduced in recent years. During the development of the Cystic Fibrosis Foundation guidelines, newborns diagnosed with CF were identified by screening through the analysis of sweat chloride levels with values ≥ 60 mmol/L³⁶.

The primary test for confirming the diagnosis of CF is the sweat chloride test, performed according to the guidelines and standards recommended by the Cystic Fibrosis Foundation committee, which places this test as the current gold standard for diagnosing CF. Moreover, it is appropriate to maintain the cut-off values recommended by the committee based on the available data from chloride testing in healthy and CF-affected infants with the following ranges for infants up to 6 months of age: ≤ 29 mmol/L, unlikely CF; 30 to 59 mmol/L, intermediate; ≥ 60 mmol/L, indicative of CF^{37,38}.

Currently, the extensive use of immunoreactive trypsin (IRT) neonatal screening has aided in diagnosing CF in asymptomatic or low symptomatic patients in approximately 64% of new diagnoses in the United States. Current guidelines on early diagnosis recommend sweat chloride testing by the Gibson and Cook technique in patients with positive screening³⁷. There is a low probability of CF diagnosis if the result is < 30 mmol/L. However, in the case of an intermediate-range (30-59 mmol/L), the test should be repeated two to three weeks later, or consider extended CFTR gene analysis or CFTR functional analysis to classify the case as CFTR-related metabolic syndrome (CRMS)/CF-screening positive, inconclusive diagnosis (CFSPID). When in both occasions the result is > 60 mmol/L, the diagnosis of CF is confirmatory, always being careful to interpret false positives (caused by conditions such as malnutrition or Down syndrome, among others) or false negatives (due to technical errors, hypoproteinemia, or sampling of the newborn in the first days). Generally, these false results are due to technical errors. Further CF functional tests, such as nasal potential difference or intestinal absorption tests, can be performed. However, these should be performed in specialized centers certified by the CF foundation³⁶.

Newborn screening for CF depends on the initial identification of elevated immunoreactive trypsinogen (IRT) values in the blood of the newborn. Since normal IRT reference values vary slightly, after identifying an abnormal IRT value, most programs perform DNA testing to detect known CFTR gene mutations (IRT/DNA strategy). This strategy provides a sensitivity of approximately 90-95% and identifies newborns at risk for a broad spectrum of disease severity³⁶.

Once the screening result is positive, sweat chloride analysis should be performed in newborns > 36 weeks of gestation and > 2 kg of weight. Genetic testing should be performed as soon as possible when available³⁸. The definitive diagnosis is based on the criteria listed in Table 2³³.

Table 2. Diagnostic criteria for cystic fibrosis

Presence of clinical criteria (at least one): Organ with consistent symptoms of cystic fibrosis Chronic sino-pulmonary disease Characteristic gastrointestinal and nutritional abnormalities Salt loss syndromes Obstructive azoospermia Sibs with cystic fibrosis Positive test result at birth (positive neonatal screen)
Tests showing CFTR dysfunction (at least one): CFTR dysfunction indicated by elevated levels in sweat chlorine analysis (> 60 mmol/L) in two tests Nasal potential difference consistent with cystic fibrosis Presence of two pathogenic variants in <i>CFTR</i> in different alleles

CF, cystic fibrosis; CFTR, CF transmembrane conductance regulator.
 Adapted from Brennan et al.³³

The diagnostic algorithm proposed in the guidelines of the CF Foundation (2017)³⁸ (Figure 3) and the one proposed by the clinical practice guideline in Mexico⁵ (Figure 4) are published and available for review.

Pulmonary function tests help assess disease severity and progression. Spirometry can be performed in children aged 3-6 years using modified acceptability criteria, and most children aged ≥ 6 years can perform reliable lung volume tests³⁹. Most patients with cystic fibrosis develop an obstructive pattern. The first manifestations of early obstruction are an increase in the ratio of residual volume to total lung capacity ratio and a decrease in forced expiratory flow of 25-75% of lung volume. Later, in advanced disease, there is a decrease in forced expiratory volume in one second (FEV1) and FEV1/FVC (forced vital capacity) ratio⁴⁰.

Chest X-rays and computed tomography are the imaging studies used in CF. Patients are usually monitored with chest X-rays every 2 years. Radiographic findings change according to the progression of the disease: hyperinflation, which later becomes persistent, can be observed. Bronchovascular markings are also visible and may progress to a pattern of bronchiectasis and cyst formation. Also, a progressive flattening of the diaphragms is present. The alterations usually appear first in the upper lobes.

CT is used when detailed knowledge of the extent of lung disease is required. High-resolution CT is useful in asymptomatic children who are small for pulmonary function testing; findings may include peribronchial thickening, air trapping, bronchiectasis, centrilobular nodules⁴¹. Pulmonary function changes may not be closely related to abnormalities found on CT (progressive).

When inconclusive results are not obtained with sweat chloride and DNA testing, the nasal potential

difference can be measured to assess CFTR dysfunction³³. The potential difference is measured by placing electrodes in the nasal cavity and comparing the voltages between baseline and administration of amiloride to block sodium transport and after nasal perfusion with a chloride-free solution, such as isoproterenol, to stimulate the CFTR-dependent chloride transporter. There is a potential difference in individuals with CFTR abnormalities at baseline, which decreases after amiloride and shows a minimal response to perfusion with chloride-free isoproterenol⁴².

Finally, a gut malabsorption study can be performed. The pancreatic exocrine function can be assessed indirectly by measuring fecal elastase. A low fecal elastase level suggests pancreatic insufficiency and consequent gut malabsorption⁴³.

Management

The management of CF is multidisciplinary and involves both the patient and family members, who will facilitate follow-up and optimization of treatment. The following are the three pillars of treatment that should be considered a guideline for all CF patients.

Keep the airway clear of secretions

Maintaining a clear airway at all times can be achieved through various techniques and schemes that can be combined according to the specific characteristics of each patient. They can be divided into two main groups:

a. Physical therapy. It seeks to clean the airways and drain the accumulation of thick, viscous secretions that can harbor bacteria and their products. Respiratory physiotherapy is performed in two or more daily sessions depending on the severity and complications, separated by a minimum of 2 hours after meals⁴⁴. It has been shown that the application of this therapy can have short-term benefits for patients with CF. However, there are as yet no conclusions on the long-term benefits⁴⁵. Techniques vary and depend on age: in children < 3 years of age, passive chest percussion maneuvers, blocks, vibrations, and postural drainage are used. After 3 years of age, forced expiration techniques can be added, inducing the child to exhale slowly and progressively, to bring secretions into the central airway. In children > 6 years, autogenic drainage maneuvers are taught³⁷.

Active cycle of breathing techniques, autogenic drainage, and Vojta approach can be combined with respiratory reeducation exercises to improve diaphragmatic function by reducing the number of respiratory cycles

Figure 3. Diagnostic algorithm for individuals with high suspicion of cystic fibrosis. CFTR, CF transmembrane conductance regulator; NPD, nasal potential difference; ICM, intestinal current measurement.

but with a higher tidal volume. In addition, physical activities that stimulate respiratory training, such as swimming, cycling, or athletics, can be implemented. One of the advantages of using specialized techniques such as physiotherapy in poorly treated patients with or patients with little or no physical activity is that it has helped to find new independent and effective ways of managing respiratory secretions. Physical exercise of different types influences the quality of life, sensitivity, and resistance to lung infections⁴⁶.

In addition to physical therapy, there are mechanical aids for the elimination of secretions. Among them, the following stand out:

- Positive expiratory pressure (PEP) mask. This mask is resistant to expiration.
 - Oscillating positive expiratory pressure (Flutter) devices. These devices combine both PEP and oscillation, generating vibrations that separate bronchial secretions throughout the airway.
 - High-frequency chest compression. This percussion vest generates pressure on the thorax through a pump, but it is costly.
 - Percussive intrapulmonary ventilator. This ventilator includes the administration of a continuous aerosol together with internal percussion of the thorax through small bursts of air at 200-300 cycles per minute⁴⁴.
- b. Pharmacological therapy

- DNase I (alpha dornase). This endonuclease breaks the DNA chains released by neutrophils responsible for the high viscosity of secretions in CF. It has been shown to improve lung function, increasing FEV1 by 6% in CF patients > 6 years and decreasing the frequency of pulmonary exacerbations. The usual dose is 2.5 mg to nebulize every 24 hours, daily and permanently, after using aerosol bronchodilators (albuterol) to prevent bronchoconstriction⁴⁷.
- Hypertonic saline solution at 7%. Pre-glucocorticoid nebulization with saline has been used to overcome hydration failure of the aqueous layer of the bronchial mucosa. This treatment reduces the frequency of exacerbations in symptomatic respiratory patients > 12 years. In children < 12 years of age, insufficient evidence has been found to justify its routine use since, in children > 6 years of age, no improvement in the frequency of exacerbations was observed. However, in tracheostomized pediatric patients, cleaning with 5% saline solution before initiating nebulizations reduces the occurrence of exacerbations³⁷. The use of inhaled hypertonic saline is suggested in young children based on randomized trials in which lung clearance index is measured and compared to placebo (isotonic saline)⁴⁸. In a randomized trial in infants < 4 months, Stahl et al. found that inhaled hypertonic saline twice daily for 52 weeks

Figure 4. Cystic fibrosis diagnosis algorithm.

NB, newborn; FMU, family medicine unit; IRT, immunoreactive trypsin.

demonstrated a significant change in lung clearance index compared to isotonic saline⁴⁹. Similar results were observed in another randomized trial in children aged 36-72 months⁵⁰.

Keep the airway free from infections

Respiratory tract infections are persistent. The administration of antibiotics, together with physical therapy and pharmacological agents that allow the elimination of secretions, will facilitate the control of the CF patient. The following is a description of the antibiotics recommended for the four main etiologic agents in respiratory tract infections:

- S. aureus* is the predominant pathogen in the first years of life and the first to infect the respiratory tract. Treatment is based on oral dicloxacillin or flucloxacillin, or intravenous dicloxacillin in exacerbations.
- P. aeruginosa* causes the most severe chronic pulmonary infection and is associated with progressive deterioration of respiratory function. Treatment consists of oral ciprofloxacin or ceftazidime, intravenous aminoglycosides or antipseudomonal penicillin, and intravenous aminoglycoside in severe cases for 21 days.
- B. cepacia* is associated with more significant pulmonary deterioration in adolescents and adults. The

incidence in Latin America is low, and treatment is based on temocillin with an aminoglycoside.

- H. influenzae* can cause chronic colonization. The indicated treatment is amoxicillin-clavulanic acid or second or third-generation cephalosporin⁵¹.

Currently, intravenous therapy has been replaced by inhaled treatments, which allow prolonged treatments with high doses and minimal toxicity. The recommended drugs are tobramycin, colistin, and aztreonam-lysine, which should subsequently be inhaled beta-blockers such as salbutamol to prevent bronchoconstriction³⁷.

Maintain an optimal nutritional state

There is a close and bidirectional correlation between body mass index (BMI) and lung function (FEV1). The goal of nutritional therapy is to maintain a BMI above the 50th percentile³⁷.

Evidence has shown that CFTR dysfunction impairs β -cells, leading to a decrease in insulin levels. Since insulin from β -cells can also exert an inhibitory effect on glucagon release from α -cells, this suggests a dual role of CFTR in regulating glucagon secretion and may contribute to glucose intolerance in CF, leading to some conditions. In the plasma of CF patients, elevated levels of peroxidized lipids and oxysterols are observed, indicating abnormal lipid metabolism and increased

susceptibility to lipoprotein lipid oxidation. In addition, pancreatic insufficiency and decreased bile acids lead to malabsorption of critical lipid-soluble antioxidants, such as carotenoids, tocopherols, and coenzyme Q-10³⁰.

The elements for adequate management of nutritional status in the patient with CF are the following: a) evaluation of BMI or the weight/height ratio in all patients; b) enzyme replacement therapy in pancreatic insufficiency; c) replacement therapy of lipid-soluble vitamins (A, D, E, K); d) hypercaloric and hyperproteic regimen to maintain metabolic goals; e) maintenance of bone health; f) replacement of bile salts in case of liver disease; g) treatment of diabetes mellitus.

Management in a specialized multidisciplinary center

It is recommended that CF patients be managed in a specialized center with the following characteristics and staff: CF specialists, a team of support specialists, a coordinating nurse, infrastructure, laboratories, and integrated outpatient care.

Children with CF regularly require hospitalization, either electively or due to acute pulmonary exacerbations or deterioration of their health status. Criteria for hospitalization are increased frequency of productive cough, increased amount or changes in the presence of sputum, increased respiratory rate, dyspnea at rest, hypoxia, marked decrease in a vesicular murmur, new changes in chest X-ray, deterioration of pulmonary function tests, weight loss or inadequate weight gain, hemoptysis⁵². As the patient progresses to advanced lung disease, options for care and prognosis should be discussed. Among these options is lung transplantation, which should be of both lungs to avoid foci of infection that could damage the transplanted lung.

The following are the referral thresholds recommended by the Cystic Fibrosis Foundation:

- In patients > 18 years:
 - a. FEV1 < 50% of the predicted value and rapidly declining
 - b. FEV1 < 40% of the predicted value and any of the following markers of shortened survival: > 2 exacerbations per year requiring intravenous antibiotics, massive hemoptysis > 240 mL requiring intensive care unit admission or bronchial artery embolization, pneumothorax, BMI < 18
 - c. FEV1 < 30% of the predicted value
- In patients < 18 years:

- a. FEV1 < 50% of the predicted value and rapidly declining
- b. FEV1 < 50% of the predicted value with any of the above markers of shortened survival or malnutrition
- c. FEV1 < 40% of the predicted value
- In all patients, any of the following, regardless of FEV1:
 - a. 6-minute walk test < 400 m
 - b. Hypoxemia (SpO2 < 88% or PaO2 < 55 mmHg) at rest or with exertion
 - c. Hypercarbia (PaCO2 > 50 mmHg) confirmed on arterial blood gases
 - d. Pulmonary artery systolic pressure > 50 mmHg on echocardiogram or evidence of right ventricular dysfunction in the absence of tricuspid regurgitant jet
 - e. Any exacerbation requiring positive pressure ventilation
- Other factors to be considered for the possibility of earlier transplantation, even when other thresholds are not met, are as follows:
 - a. Female sex, especially younger females
 - b. Short stature (height < 162 cm)
 - c. Liver cirrhosis or chronic kidney disease may require consideration of multiple-organ transplantation and may affect the timing or choice of transplant center⁵³.

Current therapies

At present, therapies to correct changes in the *CFTR* gene are focused on preventing a specific pathogenic variant from restoring messenger RNA levels (class I variants), correcting the folding and trafficking of CFTR to the apical plasma membrane (correctors for class II variants), or increasing CFTR channel function (enhancement therapy for class III variants and any variants with residual function at the apical plasma membrane). Other therapies in preclinical development are directed toward non-specific variants and include the following: gene therapy with viral and non-viral vectors (nanocarriers such as liposomes, dendrimers, exosomes); genome editing using zinc finger nuclease (ZFN) systems, effector nucleases such as transcription activators (TALENs) or CRISPR/Cas9; antisense oligonucleotides or small interfering RNAs to selectively inhibit ENaC expression; or stem cell therapy to repair airway tissue. Based on these options, personalized and more effective therapy for the CF patient can be determined^{14,54,55}.

Personalized treatment strategies include:

- a. Agents that act through reading target premature stop codon changes, such as aminoglycosides, although

they are limited due to their ototoxicity and nephrotoxicity. NB124 restored 7% of CFTR function in respiratory cell lines with G542X, R553X, R1162X, and W1282X variants; PTC124 (Atularen®), for the G442X variant, showed mixed results and no improvement, and RCT101 used nanoparticle-transported suppressor transfer RNA⁵⁶.

- b. Calcium-activated chloride channel agents (CaCCs) increase intracellular calcium release. These agents include duramycin and denufosol¹⁴.
- c. ENaC inhibitors block sodium channels used in combination with modulators. First-generation inhibitors include amiloride; second-generation inhibitors include benzamil and phenamil; third-generation inhibitors include GS-9411 and SPX-101. Camostat also indirectly inhibits ENaC by reducing Na⁺ transport through pulmonary epithelial cells¹⁴.
- d. CFTR modulators are drugs that correct protein folding, trafficking to the plasma membrane, or CFTR protein function caused by specific pathogenic variants in the *CFTR* gene. They are divided into the following groups:
 - Enhancers. They increase the probability of channel opening and enhance CFTR function, such as ATP analogs, IBMX (3-isobutyl-1-methylxanthine), genistein, PT1-808, GLPG1837, QBW251, and VX-770. VX-770 or ivacaftor prolongs the duration of channel opening, improves ion transport through protein channels making secreted mucus more fluid, decreases respiratory symptoms and the number of severe respiratory infections, increases FEV1 and weight, and decreases sweat chloride. It is authorized to treat CF caused only by specific pathogenic variants^{44,54,57,58}.
 - Correctors. They improve the folding defect, cellular processing, and trafficking of CFTR through the endoplasmic reticulum to the apical plasma membrane and increase CFTR expression on the cell surface, such as the chaperones 4-phenylbutyrate, curcumin, miglustat, and FDL169. VX-809 or lumacaftor increases chloride secretion by 14-25% for p.F508del homozygotes, but without clear clinical improvement, so some studies combine it with VX-770 (Orkambi®), although without adequate efficacy. VX-661 (tezacaftor) is also used for the p.F508del variant in homo or heterozygous state with variants with residual function, and combined with VX-770 (Symdeko®), a mean absolute improvement of 6.8% was obtained. Other correctors are currently being investigated, such as elexacaftor either alone or in double or triple combination (Trikafta®), apparently with better results in CFTR function^{44,54,59}.

- Amplifiers. These agents stimulate CFR protein expression and are applied in class V variants. They can be used in combination with enhancers and correctors such as PTI-428⁵⁴.
- Stabilizers. These molecules rectify protein instability, increase CFTR time at the plasma membrane, and decrease its degradation in the endoplasmic reticulum, such as hepatocyte growth factor, S-nitrosoglutathione (GSNO), and cavosonstat (N91115, an inhibitor of S-nitrosoglutathione reductase for p.F508del homozygotes)^{54,60,61}.

In conclusion, the measurement of IRT in the neonatal screen has allowed early diagnosis of patients with CF. However, these efforts imply challenges to homogenize diagnostic criteria and achieve more timely management, including the three pillars of treatment, to improve the quality and survival of CF patients. Research also offers new challenges and ethical issues due to its scope, but above all, a cure through precision medicine, a dream whose achievement we believe is not far off.

Ethical disclosures

Protection of human and animal subjects. The authors declare that no experiments were performed on humans or animals for this study.

Confidentiality of data. The authors declare that they have followed the protocols of their work center on the publication of patient data.

Right to privacy and informed consent. The authors have obtained the written informed consent of the patients or subjects mentioned in the article. The corresponding author has this document.

Conflicts of interest

The authors declare no conflict of interest.

Funding

None.

References

1. Comités Nacionales de Neumología, Gastroenterología, Nutrición, Grupo de Trabajo de Kinesiología. Guía de diagnóstico y tratamiento de pacientes con fibrosis quística. Actualización. Arch Argent Pediatr. 2014;112:291-92.
2. Sánchez IC, Razón BR, Ramos CLT, Barreiro PB, Reyes LC, Cantillo GH, et al. Fibrosis quística en niños y su seguimiento durante 40 años (1977-2017). Rev Cubana Pediatr. 2019;91:e882.
3. Stephenson AL, Sykes J, Stanojevic S, Quon BS, Marshall BC, Petren K, et al. Survival comparison of patients with cystic fibrosis in Canada and the United States: a population-based cohort study. Ann Intern Med. 2017;166:537-46.
4. Silva Filho LVRF, Castaños C, Ruiz HH. Cystic fibrosis in Latin America—Improving awareness. J Cyst Fibros. 2016;15:791-3.

5. Instituto Mexicano del Seguro Social. Diagnóstico de fibrosis quística en la edad pediátrica. Ciudad de México: Secretaría de Salud; 2013. Available from: <http://www.imss.gob.mx/sites/all/statics/guiasclinicas/627GRR.pdf>.
6. Goetz D, Clement L. Review of cystic fibrosis. *Pediatric Annals*. 2019;48:e154-61.
7. Scotet V, L'Hostis C, Férec C. The changing epidemiology of cystic fibrosis: incidence, survival and impact of the *CFTR* gene discovery. *Genes (Basel)*. 2020;11:589.
8. VanDevanter DR, Kahle JS, O'Sullivan AK, Sikirica S, Hodgkins PS. Cystic fibrosis in young children: a review of disease manifestation, progression, and response to early treatment. *J Cyst Fibros*. 2016;15:147-57.
9. Rey MM, Bonk MP, Hadjilidiadis D. Cystic fibrosis: emerging understanding and therapies. *Annu Rev Med*. 2018;70:197-210.
10. Guizar J. Genética Clínica. Diagnóstico y Manejo de las Enfermedades Hereditarias. México: Manual Moderno; 2001. pp. 985.
11. Sosnay PR, Siklosi KR, Van Goor F, Kaniecki K, Yu H, Sharma N, et al. Defining the disease liability of variants in the cystic fibrosis transmembrane conductance regulator gene. *Nat Genet*. 2013;45:1160-7.
12. Lay-Son G, Repetto G. Genética y fibrosis quística. Desde el gen *CFTR* a los factores modificadores. *Neumol Pediatr*. 2010;5:4-9.
13. The CFTR mutations database. Marsella: French Association Against Cystic Fibrosis. University-Hospital of Montpellier; 2013. Available from: http://www.umd.be/CFTR/W_CFTF/gene.htm.
14. Almughem FA, Aldossary AM, Tawfik EA, Alomary MN, Alharbi WS, Alshahrani MY, et al. Cystic fibrosis: overview of the current development trends and innovative therapeutic strategies. *Pharmaceutics*. 2020;12:616.
15. Molinski SV, Ahmadi S, Hung M, Bear CE. Facilitating structure-function studies of CFTR modulator sites with efficiencies in mutagenesis and functional screening. *J Biomol Screen*. 2015;20:1204-17.
16. Orozco L, Chávez M, Saldaña Y, Velázquez R, Carnevale A, González-del Ángel A, et al. Fibrosis quística: la frontera del conocimiento molecular y sus aplicaciones clínicas. *Rev Invest Clin*. 2006;58:139-52.
17. Borowitz D. CFTR, bicarbonate, and the pathophysiology of cystic fibrosis. *Pediatr Pulmonol*. 2015;50:S24-30.
18. Cantin AM, Hartl D, Konstan MW, Chmiel JF. Inflammation in cystic fibrosis lung disease: pathogenesis and therapy. *J Cyst Fibros*. 2015;14:419-30.
19. Accurso FJ, Sontag MK, Wagener JS. Complications associated with symptomatic diagnosis in infants with cystic fibrosis. *J Pediatr*. 2005;147:S37-41.
20. Sánchez I, Pérez MA, Boza ML, Lezana V, Vila MA, Repetto G, et al. Consenso nacional de fibrosis quística. *Rev Chil Pediatr*. 2001;72:356-80.
21. Jorquera P. Test de sudor en rango intermedio: desafío diagnóstico. *Neumol Pediatr*. 2016;11:15-18.
22. Suwantarant N, Rubin M, Bryan L, Tekle T, Boyle MP, Carroll KC, et al. Frequency of small colony variants and antimicrobial susceptibility of methicillin-resistant *Staphylococcus aureus* in patients with cystic fibrosis. *Diagn Microbiol Infect Dis*. 2018;90:296-99.
23. Gilligan PH. Infections in patients with cystic fibrosis: diagnostic microbiology update. *Clin Lab Med*. 2014;34:197-217.
24. Bustamante AE, Mercado-Longoria R, Tijerina-Menchaca R, Mas-Treviño M, Torres-Rodríguez J. Impacto de la erradicación de *Pseudomonas aeruginosa* sobre la sobrevida en pacientes con fibrosis quística del noreste de México. *Rev Invest Clin*. 2014;66:307-13.
25. Goss CH. Acute pulmonary exacerbations in cystic fibrosis. *Semin Respir Crit Care Med*. 2019;40:792-803.
26. Sabharwal S. Gastrointestinal manifestations of cystic fibrosis. *Gastroenterol Hepatol*. 2016;12:43-7.
27. Melo dos Santos AL, de Melo Santos H, Bettiol Nogueira M, Oshiro Távora HT, Paim da Cunha HT, de Melo Seixas RBP, et al. Cystic fibrosis: clinical phenotypes in children and adolescents. *Pediatr Gastroenterol Hepatol Nutr*. 2018;21:306-14.
28. Von Drygalski A, Biller J. Anemia in cystic fibrosis: incidence, mechanisms, and association with pulmonary function and vitamin deficiency. *Nut Clin Pract*. 2008;23:557-63.
29. O'Riordan SM, Robinson PD, Donaghue KC, Moran A. Management of cystic fibrosis related diabetes in children and adolescents. *Pediatr Diabetes*. 2009;10:43-50.
30. Kayani K, Mohammed R, Mohiaddin H. Cystic fibrosis-related diabetes. *Front Endocrinol (Lausanne)*. 2018;9:20.
31. Cano Megías M, González Albarrán O. Diabetes en la fibrosis quística: una entidad diferente. *Endocr Nutr*. 2015;62:38-44.
32. Takemoto CM. Venous thromboembolism in cystic fibrosis. *Pediatr Pulmonol*. 2012;47:105-12.
33. Brennan ML, Schrijver I. Cystic fibrosis: a review of associated phenotypes, use of molecular diagnostic approaches, genetic characteristics, progress, and dilemmas. *J Mol Diagn*. 2016;18:3-14.
34. Yoon J, Leey Casella J, Litvin M, Dobs AS. Male reproductive health in cystic fibrosis. *J Cyst Fib*. 2019;18:S105-10.
35. Hughton K, Daley T, Rayas M, Kelly A, Roe A. Female reproductive health in cystic fibrosis. *J Cyst Fib*. 2019;18:S95-104.
36. Farrell PM, White TB, Howenstine MS, Munck A, Parod RB, Rosenfeld M, et al. Diagnosis of cystic fibrosis in screened populations. *J Pediatr*. 2017;181:S33-44.
37. Fielbaum O. Manejo actual de la fibrosis quística. *Rev Med Clin Condes*. 2017;28:60-71.
38. Farrell PM, White TB, Ren CL, Hempstead SE, Accurso F, Derichs N, et al. Diagnosis of Cystic Fibrosis: Consensus Guidelines from the Cystic Fibrosis Foundation. *J Pediatr*. 2017;181:S4-S15.
39. Rosenfeld M, Allen J, Arets BHGM, Auroora P, Beydon N, Calogero C, et al. An official American Thoracic Society workshop report: optimal lung function tests for monitoring cystic fibrosis, bronchopulmonary dysplasia, and recurrent wheezing in children less than 6 years of age. *Ann Am Thorac Soc*. 2013;10:S1-S11.
40. Bergeron C, Cantin AM. Cystic fibrosis: pathophysiology of lung disease. *Semin Respir Crit Care Med*. 2019;40:715-26.
41. De Jong PA, Ottink MD, Robben SGF, Lequin MH, Hop WCJ, Hendriks JJE, et al. Pulmonary disease assessment in cystic fibrosis: comparison of ct scoring systems and value of bronchial and arterial dimension measurements. *Radiology*. 2004;231:434-9.
42. Alton EW, Currie D, Logan-Sinclair R, Warner JO, Hodson ME, Geddes DM. Nasal potential difference: a clinical diagnostic test for cystic fibrosis. *Eur Respir J*. 1990;3:922-6.
43. Daffary A, Acton J, Heubi J, Amin R. Fecal elastase-1: utility in pancreatic function in cystic fibrosis. *J Cyst Fibros*. 2006;5:71-6.
44. Romero N. Bases bioquímicas y avances en terapia de la fibrosis quística. Sevilla: Facultad de Farmacia Universidad de Sevilla; 2017. Available from: <https://idus.us.es/bitstream/handle/11441/64773/Bases%20Bioqu%EDmicas.pdf?sequence=1>.
45. Pizarro ME, Espinoza-Palma T. Tratamiento de fibrosis quística: pasado y presente. *Neumol Pediatr*. 2016;11:38-43.
46. Serrano Fernández L, Blanco-Aparicio M, Baranda García FM. La complejidad en fibrosis quística y bronquiectasias. De la multidimensionalidad en el diagnóstico a la multidisciplinariedad en el tratamiento. *Arch Bronconeumol*. 2018;5:94-107.
47. Fuchs HJ, Borowitz DS, Christiansen DH, Morris EM, Nash ML, Ramsey BW, et al. Effect of aerosolized recombinant human DNase on exacerbations of respiratory symptoms and on pulmonary function in patients with cystic fibrosis. *N Engl J Med*. 1994;331:637-42.
48. Amin R, Stanojevic S, Kane M, Webster H, Ratjen F. A randomized controlled trial to evaluate the lung clearance index as an outcome measure for early phase studies in patients with cystic fibrosis. *Resp Med*. 2016;112:59-64.
49. Stahl M, Wielpütz MO, Rieckels I, Dopfer C, Barth S, Schlegel A, et al. Preventive inhalation of hypertonic saline in infants with cystic fibrosis (PRESIS). A randomized, double-blind, controlled study. *Am J Respir Crit Care Med*. 2019;199:1238-48.
50. Felix R, Davis DS, Stanojevic S, Kronmal RA, Hinckley Stukovsky KD, Jorgensen N, et al. Inhaled hypertonic saline in preschool children with cystic fibrosis (SHIP): a multicentre, randomised, double-blind, placebo-controlled trial. *Lancet Respir Med*. 2019;7:802-9.
51. Melo J, Fernández P. Fibrosis quística en el adulto. *Rev Med Clin Condes*. 2015;26:276-84.
52. Solís-Moya A, Gutiérrez-S JP. Fibrosis quística. *Acta Med Costarric*. 2003;45:42-8.
53. Ramos KJ, Smith PJ, McKone EF, Pilewski JM, Lucy A, Hempstead SE, et al. Lung transplant referral for individuals with cystic fibrosis: Cystic Fibrosis Foundation consensus guidelines. *J Cyst Fibros*. 2019;18:321-33.
54. Pranke I, Golec A, Hinzpeter A, Edelman A, Sermet-Gaudelus I. Emerging therapeutic approaches for cystic fibrosis. From gene editing to personalized medicine. *Front Pharmacol*. 2019;10:121.
55. Bañuls L, Pellicer D, Castillo S, Navarro-García MM, Magallón M, González C, et al. Gene therapy in rare respiratory diseases: What have we learned so far? *J Clin Med*. 2020;9:2577.
56. Aslam AA, Higgins C, Sinha IP, Southern KW. Ataluren and similar compounds (specific therapies for premature termination codon class I mutations) for cystic fibrosis. *Cochrane Database Syst Rev*. 2017;1:CD012040.
57. Informe de Posicionamiento Terapéutico de Ivacaftor (Kalydeco®). Madrid: Agencia Española de Medicamentos y Productos Sanitarios. Ministerio de Sanidad, Servicios Sociales e Igualdad; 2016. Available from: <https://www.aemps.gob.es/medicamentosUsoHumano/InformesPublicos/docs/PT-ivacaftor-Kalydeco-Fibrosis-quistica.pdf>.
58. Condren ME, Bradshaw MD. Ivacaftor: a novel gene-based therapeutic approach for cystic fibrosis. *J Pediatr Pharmacol Ther*. 2013;18:8-13.
59. Verkman AS, Galletta LJ. Chloride channels as drug targets. *Nat Rev Drug Discov*. 2009;8:153-71.
60. Donaldson SH, Solomon GM, Zeitlin PL, Flume PA, Casey A, McCoy K, et al. Pharmacokinetics and safety of cavaconstat (N91115) in healthy and cystic fibrosis adults homozygous for F508DEL-CFTR. *J Cyst Fibros*. 2017;16:371-9.
61. Bhagirath AY, Li Y, Somayajula D, Dadashi M, Badr S, Duan K. Cystic fibrosis lung environment and *Pseudomonas aeruginosa* infection. *BC Pulm Med*. 2016;16:174.